# Why is antimicrobial resistance in pigs not so important for public health?

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#### **Abstract**

In recent years there has been growing concern over the potential spread of antimicrobial resistance from the use of antibiotics in animals to humans and that they may play a significant role in causing resistance in man. Lay press have expressed up to 70% of resistance seen in man may come from the agricultural use of antibiotics, even risk assessment reports to governments say there is no hard evidence but an approximate estimate of 33-50% of resistance may come from the agricultural sector. It was the purpose of this paper to evaluate the potential risk of the transmission of infections from pigs to man either directly or via pig meat using an epidemiological basis. This could then be compared with the incidence of diseases in man caused mainly by bacteria and are likely to be treated with antibiotics. Data evaluation showed that pigs could be accountable for infections in man, especially personnel working directly with pigs. Regarding the general public, the figure was much lower at 0.0031% on a population basis, which is equivalent to 3.1 people in every 100,000. Human infections occur and are treated with antibiotics in approximately 16.34% of cases on a population basis, or 16,340people/100,000. Therefore pigs cause 0.019% of human cases and if all infections were resistant to antimicrobials this would be the contribution of resistance transmission. If the concern was just for 'critical use' antibiotics the figure would be as low as 0.00038% if a figure of 2% resistance is used. In conclusion, this is the reason why antimicrobial resistance transfer to the general human population from pigs can be considered of minor importance in comparison with the direct use of antimicrobials in man by doctors in the community and in hospitals.

# Introduction

Antibiotics have been extensively used in swine production over many years. There have been recent reports of outbreaks of methicillin-resistant *Staphylococcus aureus* (MRSA) CC398 in pigs starting in the Netherlands (Voss et al, 2005) and now recognised across most of the European Union (EFSA 2009, 2010). There have been increasing reports of the presence of extended-spectrum beta-lactamase (ESBL) producing bacteria, especially *Escherichia coli* and *Salmonella* spp (EFSA, 2011a), associated with the increasing use of 3<sup>rd</sup> and 4<sup>th</sup> generation cephalosporins. Fluoroquinolone resistance has also been reported to be on the increase, especially in poultry (MARAN, 2011). As a result some countries, such as the Netherlands, have decided to reduce their antibiotic consumption in veterinary medicine by 50% by 2013. The European Commission (2011) and the European Council (2012) have outlined the review of antimicrobials in the EU over the next 5 years and the details are awaited with keen interest.

However, much of the views regarding antimicrobial resistance transfer from animals to man is speculative and it is difficult to obtain finite information of the actual risks. However, it does not stop the lay press expressing opinions of the quantity and a figure of up to **70%** has been promulgated. Even in more sophisticated reports (BuRO, 2010) the Bureau of Risk Assessment and Research Programming in a report to the Dutch Government stated, "There are no hard evidence on the actual importance of transfer of resistance from the animal to the human sector but a rough estimate is that **33-50% of the resistance** observed in infectious agents for humans, comes from the agricultural sector."

The purpose of this paper is to try to quantify the transmission of bacterial infections from pigs to man and the potential transfer of antimicrobial resistance, using an epidemiological approach.

The epidemiological approach – the chain of infection

It is necessary to identify the infectious agent to be evaluated; the source is the pig, the route of transmission to the host, man, the susceptibility of the host to the colonisation and infection regarding a zoonotic bacterium. If a commensal bacterium, then the rate of colonisation and resistance transmission from the swine bacterium to the host bacterial flora needs to be determined if possible. A further step for commensal bacteria would be to evaluate the significance of those bacteria in the host regarding them causing infection (often associated with nosocomial/hospital infections) and their resistance affecting drug treatment responses.

Table 1. The chain of infection and resistance for zoonotic and commensal bacteria (Prescott et al 2005)

The chain	Zoonotic infection	Commensal infection
Organism	Bacteria	Bacteria
Source	Pig	Pig
Host	Man	Man
Route	Direct, indirect, food etc	Direct, indirect, food etc
Susceptibility of host	Natural defences, age, immune	Natural defences, age, immune
	status	status
Colonisation	Yes	Yes
Infection	Yes	No (Yes – nosocomial?)
Disease incidence (%)	Yes	No (Yes - nosocomial?)
Resistance transfer	Yes	Yes
Mortality incidence	Yes	No (Yes – nosocomial?)
Treatment failure	Yes	Yes
incidence		

### **Human bacterial infections**

A list of human bacterial infections and their incidence (mainly US data) are summarised below (see Table 2) and the potential associations with swine identified (Prescott et al, 2005; Mossad, 2010; Foxman, 2002).

Table 2. Human bacterial infections and potential swine association (Prescott et al, 2005; Mossad, 2010; Foxman, 2002)

Human bacterial infections	Information - incidence	Potentially swine associated
Airborne / respiratory	39 million upper respiratory tract infections (URTI)	
	cases/year US; 75% receive antibiotic prescriptions	
Lower respiratory tract infection	ons (LRTI) (pneumonias)	
Corynebacterium diphtheriae	<100 cases in US/year	
Legionella pneumophila	1000-1400 cases/year US; 30,000 undiagnosed	
Bordetella pertussis	5,000 cases, 10 deaths/year US; 500,000 deaths WW	
Streptococcus pyogenes	Man asymptomatic carrier	
Streptococcus pneumoniae	Asymptomatic carrier; 60-80% of pneumonias; 150-	
	300,000 cases, 13-66,000 deaths in US	
Haemophilus influenzae		
Mycobacterium tuberculosis	WW 1 billion infected, 10 million cases/year, 3	

Human bacterial infections	Information - incidence	Potentially swine associated
	million deaths; US 26,000 cases, 12,000 deaths/year	
Mycoplasma pneumoniae	Mortality <1%	
Chlamydia pneumoniae	·	
Meningitis		
Streptococcus pneumoniae		
Neisseria meningitidis		
Haemophilus influenzae		
Group B streptococci		
Listeria monocytogenes		
Mycobacterium tuberculosis		
Nocardia asteroides		
Staphylococcus aureus		Yes
Staphylococcus epidermidis		
Arthropod-borne infections		
Borrelia burgdorferi	20,000 cases/year US	
Yersinia pestis	25 cases 15% deaths US	
Direct contact diseases		
Bacillus anthracis	<1 case/year US; 20-100,000 cases WW	Yes
Clostridial infections		
Mycobacterium leprae	4000 cases 2-300 new/year US; 14million WW	
Sexually transmitted	8-10 million/year US	
Neisseria gonorrhoeae	Most common US	
Treponema pallidum		
Gardnerella vaginalis		
Haemophilus ducreyi		
Chlamydia trachomatis		
Mycoplasma genitalium		
Mycoplasma hominis	Widespread	
Ureaplasma urealyticum	Widespread	
Urinary tract infections (UTI)	8 million case/year US; women 14 times more likely	
	than men; >1 million catheter-associated UTI in	
	hospitals and care homes	
Food and water borne		
Staphylococcus aureus		Yes
Bacillus cereus		
Clostridium perfringens		Yes
Clostridium difficile		Yes
Clostridium botulinum	<100 cases/year US	
Escherichia coli		Yes
Vibrio cholerae	<20 cases/year, <1% deaths US; 600,000 deaths/year WW	
Vibrio parahaemolyticus		
Shigella	25-30,000 cases/year US; 600,000 deaths/year WW	
Salmonella typhi	4-500 cases/year, <1% mortality US	
Salmonella spp	45,000 cases reported/year US; suspect 2-3million	Yes
Campylobacter jejuni	2 million case/year US	
Campylobacter coli	10% human cases	Yes

Human bacterial infections	Information - incidence	Potentially swine associated
Yersinia enterocolitica		Yes
Listeria monocytogenes		
Nosocomial (hospital)		
Staphylococcus aureus	LRT, surgical wounds, skin, bacteraemia	Yes
Escherichia coli	UTI, surgical wounds, skin,	Yes
Pseudomonas aeruginosa	UTI, LRT, surgical wounds, skin, bacteraemia	
Enterococcus spp	UTI, surgical wounds, bacteraemia	Yes
Enterobacter spp	UTI, LRT, surgical wounds, bacteraemia	
Acinetobacter spp	LRT	
Klebsiella pneumoniae	LRT	

Key: US = United States; WW = Worldwide; URTI = upper respiratory tract infection; LRT = lower respiratory tract infection; UTI = urinary tract infection

Of the 49 infectious agents reported, potentially only nine are pig associated (approximately 18%) but they may also involve other animal species. A further bacterium, *Streptococcus suis* is also potentially transmissible to man from pigs, but usually affects farm workers or persons involved in the meat trade.

# Major infections that are potentially associated with pigs

An overview of the percentage incidence of infection and mortality of some human and potentially pig associated diseases are summarised in Table 3.

Table 3. The percentage incidence of infection and mortality of some human diseases on a population basis

Infection	Incidence (population %)	Mortality (cases %)	Pig associated
Respiratory			
URTI	13 pa (75% treated = 9.75) (US)		
LRTI (pneumonias)	0.13 (US)		
S. pneumoniae	0.05-0.1 pa (US)	8.7-22.0 (US)	
Sexually transmitted	2.6-3.3 pa (US)		
Urinary tract infections	2.7 (US)		
(UTI)			
Food & water borne	0.76 (US)		
S. typhi	0.0017 pa (US)	<1.0 (US)	
Salmonella spp	0.015 reported; est 1.0 pa (US)		Yes
C. jejuni	0.67 pa (US)	0.005pa (US)	
C. coli	0.07 pa (US)		Yes

Key: Populations – US 300 million; pa – per annum

If the major bacterial infections that are potentially associated with pig transmission are examined further then their impact regarding infection, disease, mortality and resistance transfer can be further assessed (see Table 4).

Table 4. Comparison of the likelihood of bacterial agents causing disease and causing resistance issues

Bacterium	Colonisation	Infection	Disease	Death	Resistance
S. aureus (MRSA CC398)	Yes (DK)	Yes (DK)	Yes (DK)	?	Yes
	No (UK)	No (UK)	No (UK)	No	No
S. suis	Yes	Yes	Yes	Yes	No (penicillin Sens)
B. anthracis	Poss?	Poss?	Poss?	Poss?	No (penicillin Sens)
C. perfringens	Poss?	Poss?	Poss?	Poss?	No (penicillin Sens)
C. difficile	Poss?	Poss?	Poss?	Poss?	Poss? (nosocomial)
A. baumannii	Poss?	Poss?	Poss?	Poss?	Poss? (nosocomial)
E. coli	Poss?	Poss?	Poss?	Poss?	Poss? (nosocomial)
Salmonella spp	Yes	Yes	Yes	Yes?	Yes
C. coli	Poss?	Poss?	Poss?	Poss?	Poss?
Enterococcus spp	Poss?	Poss?	Poss?	Poss?	Poss?
Y. enterocolitica	Poss?	Poss?	Poss?	Poss?	Poss?

Key: Poss? – possible; penicillin Sens – penicillin sensitive

### Streptococcus suis, Bacillus anthracis, Clostridium perfringens

Streptococcus suis has caused disease in man (Barlow et al, 2003) and in the UK **21**% of stockmen were found to be seropositive. There were approximately **2 cases/year** over a 20 year period recording period, with cerebrospinal fluid or blood isolates and with **12.5**% of cases dying from their infection. The majority of cases showed signs of meningitis and septicaemia and involved mainly pig farmers/stockmen, abattoir workers and butchers (93% of cases with epidemiological data). Resistance was not an issue as most *S. suis* isolates were considered penicillin susceptible in the UK.

Bacillus anthracis is uncommonly found in pigs in the UK and could possibly infect man, closely involved with an outbreak but the majority of isolates are considered penicillin sensitive. Clostridium perfringens could also potentially infect man but there is little evidence of disease. Resistance is not an issue as the majority of pig isolates are susceptible to penicillin.

# Methicillin resistant Staphylococcus aureus (MRSA)

In contrast to *S. suis*, MRSA strains have been a major problem in man especially associated with hospital infections but also increasing in the community. The MRSA clone CC398 has been found in many EU countries in pigs and now is being found in man, mainly in countries which have a high pig farm MRSA prevalence e.g. Netherlands, Germany, Spain and Italy. Pig farmers in Germany can be highly infected (83%) but generally the infection has remained in farmers, their family members (4.3%) and that it has not spread to any major extent into the general public in contact such as schools that were monitored (Cuny et al, 2009). Swine veterinarians (36%), their laboratory staff and abattoir workers (14%) were also shown to have a higher incidence of infection than the national average (1-2%) in Germany (Blaha et al, 2009). In Denmark, there has been an increase in cases of MRSA in man. Cases here refer to colonisation and clinical infections (see Figure 1).

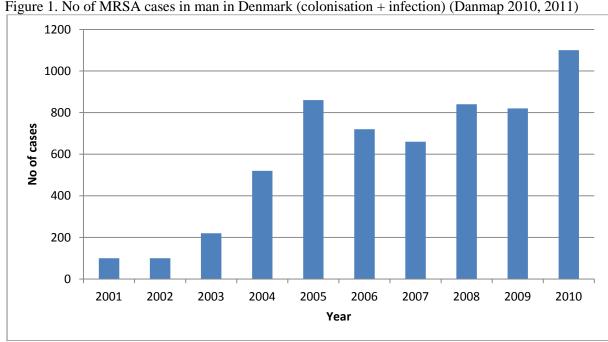


Figure 1. No of MRSA cases in man in Denmark (colonisation + infection) (Danmap 2010, 2011)

Approximately, 4.95% of 646 human MRSA clinical infections were associated with CC398, the livestock associated (LA) clone in 2010 (see Figure 2). Asymptomatic carriers were higher at 13.53% of 451 isolates (Danmap 2010, 2011).

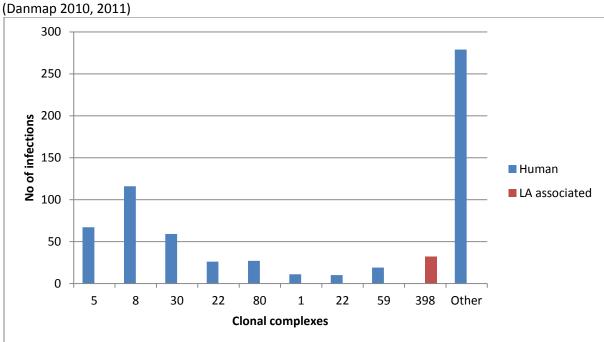


Figure 2. Common clonal complexes of MRSA causing clinical infections in man in Denmark in 2010

Overall, MRSA CC398 played a 9.57% role in a rapidly growing problem in Denmark, where MRSA new cases (both infected and only colonised) increased from approximately 100 in 2001 to 1097 in 2010 (11-fold increase). This has led to a voluntary suspension of use of 3<sup>rd</sup> and 4<sup>th</sup> generation cephalosporins in veterinary medicine in 2010 and the implementation of a number of other

controls to reduce antimicrobial use. On a population basis (5.5 million), MRSA cases were **0.02%** in Denmark and LA MRSA cases account for **0.002%**.

In spite of the UK being free of LA MRSA in pigs, its Health Protection Agency (HPA) reported that it has gone through a major MRSA epidemic but this was linked primarily to the medical arena and poor hospital hygiene practices (see Figure 2).

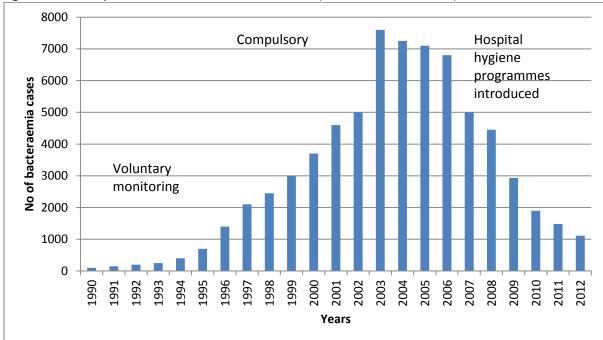


Figure 2. MRSA epidemic of bacteraemia cases in UK (HPA, 2006; HPA, 2012)

The introduction of hospital hygiene initiatives has resulted in a dramatic fall (-85%) in the incidence of MRSA bacteraemia cases in the UK from **0.012**% of population down to **0.0018**%. This highlights the major nosocomial impact of clinical MRSA infections in man, in comparison with LA MRSA associated transmission.

# Zoonotic and food-borne outbreaks (EFSA, 2011b)

EFSA (2011b) reported on the incidence of confirmed cases of various zoonotic and food-borne infections in humans in the EU from 2005-09.

(EFSA, 2011b) 220 200 180 160 Confirmed cases ('000s) 140 Campylobacter 120 Salmonellosis 100 Yersiniosis 80 E.coli VTEC 60 40 20 0 2007 2008 2005 2006 2009

Figure 3. Incidence of confirmed cases of zoonotic and food-borne infections in humans 2005-09

Campylobacter infections have remained fairly steady over the 5-year period but there has been a steady fall in Salmonella cases. *Yersinia* spp and *E. coli* VTEC cases were comparatively low by comparison and Yersinia infections will not be dealt with further.

# **Campylobacter** spp infections

Burch (2002) reviewed the likely transmission of *C. coli* from pigs to man. The majority of isolates in man was *C. jejuni* (92%) and *C. coli* (8%). This was in contrast to the findings in pigs (see Table 5).

Table 3. The isolation incluence (70) of Campylobacter spp from numaris and animals (burch, 2002	Table 5. The isolation incidence (	6) of Campylobacter spp from humans and anima	Is (Burch, 2002)
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Species	C. jejuni (%)	C. coli (%)
Humans	92	8
Chicken	90	10
Cattle	99	1
Pigs	4	96

Generally, it was assumed that chickens/chicken meat were the main route of transmission of *C. jejuni* to man, as there was a high incidence of carcass contamination but pigs were still considered the main source of *C. coli*. EFSA (2011a) showed that 31% of broiler carcasses and meat were contaminated with *Campylobacter* spp but only 0.6% of pig meat. In addition, the species relationships in man were in complete contrast and were similar to chicken isolates.

At that time (Burch, 2002), the macrolide tylosin was still extensively used in pigs and there was a relatively high level of resistance reported to erythromycin, another macrolide. The erythromycin resistance pattern was then compared to see whether a link or assessment of transmission of *C. coli* to humans could be estimated (see Table 6).

Table 6. Comparison of erythromycin (macrolide) resistance in Campylobacter spp from different animal sources (Burch, 2002)

Species	C. jejuni (%)	C. coli (%)
Humans	2	15
Chicken	4	15
Cattle	1	-
Pigs	35	57

As the *C. coli* erythromycin-resistance profiles in pigs were completely different to those found in humans but were almost identical to chickens, it was considered that the contribution of infection from *C. coli* in pigs was probably very low, possibly **zero**.

This assumes that all infections are meat transmitted. The main incidence of campylobacteriosis in man increases in the summer months, so it was thought that barbecues might have an impact. Other sources might also contribute, such as swimming and beach holidays. In the US, wildfowl such as Canadian geese (52%) were excretors of *C. jejuni* as well as Mallard ducks (40%) (Fallacra et al, 2001), so fresh water swimming may also be an issue. Picnicking near cattle might also be a problem, as well as beach holidays. Dingle et al (2002) showed that *C. jejuni* found in beach sand was identical to human strains. Could this arise from sewage release into the sea?

## Salmonella spp infections

In the UK, there have been many changes in the incidence of Salmonella infections in man over the last decade (VLA, 2002; VLA, 2007; AHVLA, 2012).

Table 7. Main Salmonella serovars found in humans in the UK between 2001-2011

Year	2001	2006	2011
No of cases reported	18036	13412 (-25.6%)	8937 (-50.4%)
Main serovar			
Enteritidis	65.1	55.3	30.4
Typhimurium*	13.0	12.1	25.9 (*+monophasic)
Virchov	2.2	3.2	1.5
Hadar	1.5	<0.7	<1.0
Newport	1.0	1.8	2.0
Infantis	1.0	1.2	1.9
Braenderup	0.9	0.7	<1.0
Agona	0.8	<0.7	1.6
Paratyphi B var Java	0.7	<0.7	1.3
Stanley	0.6	1.2	<1.0
Others	13.2	23.8	35.2

The number of reported human cases has fallen by 50.4%, a substantial fall. There has been a dramatic fall in cases of *S*. Enteritidis from 11747 to 2720 (-76.8%). *Salmonella* Enteritidis was primarily found in chickens and eggs and with EFSA's and the poultry industries initiatives to reduce the incidence in poultry by vaccination of breeding hens and monitoring of flocks this has fallen substantially. It must be remembered that cases picked up while travelling abroad are also included. In contrast, *S*. Typhimurium the more pig associated bacterium has increased in percentage terms but in case terms has stayed approximately the same at 2340 in 2001 and 2315 in 2011. On this basis, the incidence of Salmonella infections in reported cases in humans in the UK is **0.014%**, S. Enteritidis is **0.0043%** and the incidence of *S*. Typhimurium cases is **0.0037%**.

Table 8. Comparison of the major *Salmonella* serovars found in humans and animals in 2011 (AHVLA, 2012)

Human	Cases (%)	Pigs	Cases (%)	Chickens	Cases (%)	Cattle	Cases (%)
Enteritidis	30.4	Typhimurium	42.3	Montevideo	21.0	Dublin	65.0
Typhimurium*	25.9	4,5,12:i:-	22.0	Kedougou	19.3	Mbandaka	12.9
Newport	2.0	4,12,:i:-	11.0	Senftenberg	11.7	Typhimurium	7.3
Infantis	1.9	Derby	7.1	Livingstone	10.2	4,5,12:i:-	3.8
Montevideo	1.9	Bovismorbificans	3.3	Mbandaka	9.5	Montevideo	3.5
Agona	1.6	Kedougou	2.2	Ohio	7.1	Anatum	1.7
Kentucky	1.5	London	2.2	Typhimurium	2.9	Agama	0.8
Virchow	1.5	Newport	2.2	Enteritidis	2.0	Enteritidis	0.8
Paratyphi B	1.3	Panama	2.2	Derby	1.5	4,12:i:-	0.6
var Java							
Oranienburg	1.1			Thompson	1.2	Infantis	0.4
Others	30.7	Others	5.5	Others	13.7	Others	3.1

<sup>\*</sup>Typhimurium + monophasics; Pigs = 75.3% isolates; Cattle = 11.7%

It is very hard to apportion responsibility for different species of animal's serovars that contribute to the incidence of disease in man. It is unfortunate that the incidence of monophasics in man is not recorded separately, as this would also add to the epidemiological knowledge of transmission. Pigs do carry a lot of *S.* Typhimurium (75.3% of isolates) and therefore has always been considered responsible for the bulk of transmission, yet poultry and cattle are also carriers.

The AHVLA (2012) further subdivided the S. Typhimurium serovars into phage types (PTs)

Table 9. Phage types (PT) of *S.* Typhimurium from humans and different animal species (AHVLA, 2012)

Human	PT (%)	Pigs	PT (%)	Chickens	PT (%)	Cattle	PT (%)
DT193	28.6	U288	37.7	DT40	16.7	DT104	32.7
DT120	14.1	DT193	22.1	DT193	16.7	DT193	19.2
DT104	7.5	DT120	14.3	UNTY	16.7	DT120	11.5
DT8	5.4	U302	7.8			U302	9.6
RDNC	4.1	UNTY	6.5			DT8	5.8
U320	2.9	DT104B	3.9			DT2	3.8
DT191a	2.4					DT12	3.8
DT104b	2.1					U289	3.8
U323	1.9					UNTY	3.8
Others	31	Others	7.8	Others	50	Others	5.8
Human top 4 total	55.6		36.4		16.7		69.2
Est. proportional contribution (%)	55.6		28.4		8.2		28.7

It is possible that pigs only account for **28.4%** of the major *S.* Typhimurium infections in man or **7.4%** of Salmonella cases or **0.0011%** on a population basis in 2011.

The resistance profiles were examined for critically important antibiotics like the fluoroquinolones and 3<sup>rd</sup> and 4<sup>th</sup> generation cephalosporins in the UK. There was no resistance to cefotaxime or ceftazidime (both 3G cephalosporins) or amoxicillin/clavulanic acid by *S*. Typhimurium but there was

surprisingly in 4 isolates of *S.* Derby. There was no ciprofloxacin resistance or amikacin resistance but there was 23% apramycin/gentamicin resistance mainly in the new monophasic Salmonella. Resistance to the 'critical use' antimicrobials is relatively low in the UK and therefore treatment failures due to resistance are also likely to be very low.

In Europe (EFSA, 2011c) reported on the comparative resistance of *S.* Typhimurium isolates from humans, pigs, cattle and poultry in 2009 (see Figure 4.)

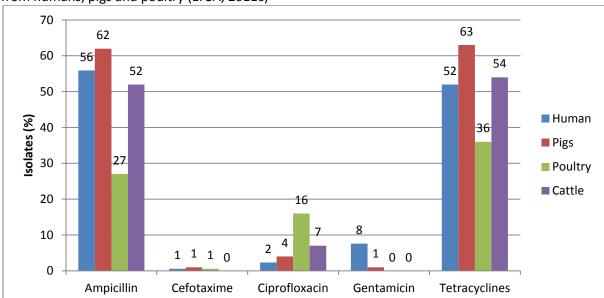


Figure 4. Comparative EU resistance profiles of various antimicrobials in *S.* Typhimurium isolates from humans, pigs and poultry (EFSA, 2011c)

There is some variation in the resistance profiles between animal and human species. None fit precisely but pig and cattle are more similar than poultry, except for gentamicin, where human resistance is comparatively high. Critical antibiotic resistance, say the fluoroquinolones (4%), from porcine *S.* Typhimurium might only account for **0.000044**% of resistance on a population basis and 3G cephalosporins (1%) even less at **0.000011**%.

## Escherichia coli infections

From a public health point of view, *E. coli* VTEC isolates are the most significant with O157 serotype accounting for 51.7% of strains in 2009 (EFSA, 2011c). The main sources identified were mainly from meat from cattle (2.3% of samples) raw milk and sheep meat (3.2%). There is a high summer incidence and children are most commonly affected. Pig meat does not appear to be involved.

Transmission of *E. coli* to pig farmers was investigated by Nijsten et al (1996) in the Netherlands. They found that antibiotic resistance of *E. coli* isolates in faecal samples of pig farmers was significantly lower than samples obtained from pigs. The resistance patterns of only **4%** of farmer *E. coli* were the same as pigs from the same farm. In many ways, this is surprising, as one would have expected a high direct exposure over a prolonged period, although via the faecal/oral route, and would have caused a much higher colonisation/contamination rate, especially in comparison with the high MRSA colonisation rate seen in German farmers via the nasal/respiratory route.

This suggests that pig *E. coli* may not be able to colonise the intact human gut that well, with all its defence mechanisms (e.g. stomach acid, competitive and host-adapted gut flora). The result of failed colonisation did not appear to contribute greatly to antimicrobial resistance either. Any impact

on pig-meat consumers can be considered much less in comparison with pig farmers but this has not been quantified.

Table 10. Comparison of pig and pig farmer *E. coli* antimicrobial resistance profiles (Nijsten et al, 1996)

Antimicrobial	Pig resistance (%)	Pig farmer resistance (%)
Amoxicillin	25	28
Amoxicillin/clavulanate	0	0
Apramycin	0	0
Chloramphenicol	13	7*
Nitrofurantoin	8	3*
Nalidixic acid	0	2
Neomycin	7	3
Oxytetracycline	57	32*
Streptomycin	71	35*
Sulphamethoxazole	45	35*
Trimethoprim	16	10

Key: \* = p = < 0.05

In VMD (2010) they compared the antimicrobial susceptibility of UK *E. coli* causing bacteraemia in humans with clinical submissions for pigs and chickens.

Table 11. Comparison of human and animal clinical *E. coli* isolates and their resistance (%) (VMD, 2010)

Antimicrobial	Human (n=>10,000 isolates)	Pig (n=73-231 isolates)	Chicken (n=68-71 isolates)
Ampicillin	61	46	44
Amoxycillin/	20	1	-
clavulanic acid			
Cefpodoxime (3 <sup>rd</sup> gen)	-	1	1
Ceftazidime (3 <sup>rd</sup> gen)	11	-	-
Cefotaxime (3 <sup>rd</sup> gen)	12	-	-
Enrofloxacin/		6	6
Ciprofloxacin	23		
Neomycin	-	9	3
Apramycin	-	10	1
Gentamicin	9	-	-
Tetracycline	-	78	51
Trimethoprim	40	53	24
(sulpha)			
Piperacillin/ Tazobactam	8	-	-

The first observation is the difference in antimicrobial drugs that are used for testing between human and animal use. Ampicillin/amoxycillin demonstrate high resistance in both humans and animals, but when combined with the beta-lactamase inhibitor, clavulanic acid, the resistance falls dramatically in animals but not in humans, where there appears to be a 20% resistance. All the cephalosporins are 3<sup>rd</sup> generation and again it demonstrates a very low level of resistance in pigs and chickens in comparison with human isolates. Not all of these resistant isolates will be carrying the ESBL resistant genes but they have been found in 9.2% of human cephalosporin-resistant *E. coli* and in animals mainly in cattle isolates but once in chicken isolates. Fluoroquinolone resistance is also

lower in animal medicine in comparison with human medicine. Aminoglycoside resistance is similar in both man and pigs at about 9-10% and trimethoprim (+ sulphas) are also high in man and pigs in comparison with chickens.

Overall, the data suggests that the transference of *E. coli* to consumers by meat is relatively low, the transfer of resistance can also be considered low and the impact on the use of antibiotics in human medicine has the main impact on human antimicrobial resistance.

# **Enterococcus** spp infections

It is difficult to obtain precise data about the colonisation of the human gut by Enterococci from animals. Danmap 2006 (2007) offers some insight as they compared the resistance profiles of *E. faecium* and *E. faecalis* from pigs, broilers and turkeys and healthy humans.

Table 12. Comparison of resistance profiles in *E. faecium* from healthy humans pigs and broilers (Danmap, 2007)

Antimicrobial	Humans	Pigs	Broilers
Tetracycline	8	61	7
Ampicillin	4	0	0
Erythromycin	8	34	29
Gentamicin	0	0	0
Kanamycin	13	25	0
Streptomycin	4	30	14
Vancomycin	0	3	0*
Quinupristin/dalfopristin	0	1	1

Key: \*Using selective media vancomycin resistant *E. faecium* was found in 47% of broiler samples but not in pig and cattle (Danmap, 2011)

The resistance profiles of *E. faecium* in humans were very different to those found in pigs and broilers, especially with regard to tetracycline, ampicillin (converse), erythromycin, kanamycin and streptomycin. A maximum of **13%** using tetracycline could be transmitted but unfortunately, the precise profiles for each isolate were not compared.

The differences in *E. faecalis* resistance profiles were less marked, but again they were quite different. Individual isolate profiles were not compared thus a likely percentage figure could not be determined.

Table 13. Comparison of resistance profiles in *E. faecalis* from healthy humans pigs and broilers (Danmap, 2007)

(			
Antimicrobial	Humans	Pigs	Broilers
Tetracycline	39	85	27
Ampicillin	0	0	0
Erythromycin	7	38	20
Gentamicin	0	4*	0
Kanamycin	10	23	0
Streptomycin	10	32	0
Vancomycin	0	0	0

Key: \*Potential reservoir/association with human gentamicin-resistant endocarditis MLST type ST16 (Danmap, 2011) or primarily nosocomial/hospital problem not determined.

Considering that the bacterium needs to be transmitted via pig meat, which is sometimes eaten raw in Denmark but usually cooked or cured, the resistance profile would suggest a low colonisation rate and low resistance transfer rate to healthy humans of **18%** maximum from pigs, using erythromycin as a marker. Individual isolate resistance profile was not compared.

#### Other bacteria

With other bacteria, such as *C. difficile*, the types found in animals (pigs, cattle and broilers) and human clinical cases could not be directly compared since the human isolates were selected for typing based on the O27 ribotype by PCR and also moxifloxacin resistance (Danmap, 2011). The O78 ribotype, which has been found in human cases, was found in cattle and pig isolates but were moxifloxacin sensitive. *Acinetobacter baumanii* isolates from human nosocomial infections were shown not to be derived from animal isolates using PFGE profiles (Hamouda et al, 2011).

#### **Conclusions**

If one compares the estimated disease incidence caused by pigs, transmitted to man, it can be seen to be relatively small and can be calculated as **0.003103%** on a population basis (see Table 14).

Table 14. Overview of potential infection transmission to humans from pigs

	Zoonotic infections			
Organism	S. suis	MRSA	C. coli	S. Typhimurium
Source	Pig	Pig	Pig	Pig
Host	Farmer/Man	Farmer/Man	Man	Man
Route	Respiratory/wound	Respiratory	Food/meat	Food/meat
Host susceptibility	Moderate	High	High	High
Colonisation	Low, 21% Farmer/ V. low man	High >83% Farmer/ 9.57% cases/ V. low man	No	Yes 25.9% cases
Infection	Very low	Moderate 4.94%	No	Moderate 7.4%
Disease incidence	0.00003%	0.002%	0%	0.0011%
Resistance transfer	No	No	No	Yes, potential
Mortality incidence	12.5%	No	No	Low
Treatment failure incidence	No	No	No	Low
	Commensal infections			
Organism	E. coli	Enterococci	C. difficile	A. baumannii
Source	Pig	Pig	Pig	Pig
Host	Farmer/Man	Man	Man	Man
Route	F/O; Food/meat	Food/meat	Food/meat	Food/meat
Host susceptibility	Low, 4% Farmer	Undefined	Undefined	Undefined
Colonisation	Low undefined	Undefined	Undefined	Undefined
Infection	No (commensal)	No (commensal)	No (nosocomial)	No (nosocomial)
Disease incidence	No (commensal)	No (commensal)	No	No
Resistance transfer	Yes potential	Yes potential	No	No
Mortality incidence	No (commensal)	No (commensal)	No	No
Treatment failure incidence	Very low undefined	Very low undefined	No	No

Key: F/O = Faecal/oral

If the disease incidence caused by pigs is compared with infections in man, which are likely to be treated with antibiotics and in their own right select for resistance the proportion due to use in pigs can be considered minimal almost inconsequential (see Table 15).

Table 15. Relative case and resistance transmission from pigs to humans

Pig transmission	Transmission (%) on a	Human infections	Population affected
	population basis		and treated (%)
S. suis	0.000003	Respiratory (URTI &	9.88
		LRTI)	
MRSA	0.002	Sexually transmitted	2.6-3.3 (say <b>3.0</b> )
Campylobacter spp	0.0	Urinary tract	2.7
S. Typhimurium	0.0011	Food and water borne	0.76
Total	0.003103	Total	16.34
	Transmission on a case		
	basis		
Cases/100,000	3.1		16,340
population			
Case potentially	0.019%		100%
transmitted from pigs			
to man			
Resistance	0.00038%		100%
transmission of critical			
antibiotics (at 2%)			

If the disease incidence in humans associated with bacteria from pigs at **0.003103%** on a population basis is compared with human infections of **16.34%** on a population basis per year, which are likely to be treated with antimicrobials, the percentage of infections potentially caused by pigs is approximately **0.019%** of human infections. If all the bacteria coming from pig infections were resistant, it would only contribute **0.019%** of resistance to humans. The resistance rate of 'critically important' antimicrobials is about **2% (0-6%)**, then contribution to human resistance of key antibiotics is likely to be about **0.00038% (0.0-0.002%)**, which is a remarkably small amount. That is why antimicrobial resistance in pigs is not so important for public health, in comparison with the amount of resistance that human doctors directly produce, especially in the hospital situation.

### **References**

AHVLA (2012) Salmonella in livestock production in GB, 2011. <a href="http://vla.defra.gov.uk/reports/rep\_salm\_rep11.htm">http://vla.defra.gov.uk/reports/rep\_salm\_rep11.htm</a>

Barlow, A.M., Hunt, B.W., Heath, P.J. and Smith, R.M.M. (2003) The prevalence and clinical diseases caused in pigs by different serotypes of Streptococcus suis (June 200-September 2002) and human infection (1981 to October 2002) in England and Wales. Pig Journal, 51, 164-176.

Blaha, T., Eckmans, T., Cuny, C., Witte, W. And Meemken, D. (2009) Methicillin-resistant Staphylococcus aureus (MRSA) in veterinarians and meat inspectors. Proceedings of the 14<sup>th</sup> International Society for Animal Hygiene (ISAH) Congress, Vechta, Germany, vol 2, pp 645-648.

Burch, D.G.S. (2002) Risk assessment – Campylobacter infection transmission from pigs to man using erythromycin resistance as a marker. Pig Journal, 50, 53-58.

BuRo (2010) Bureau of Risk Assessment and Research Programming (BuRo) report – Advice on increased resistance due to concentration of AB (antibiotics) after CO (carry-over). Dated 10/12/2010.

Cuny, C., Nathaus, R., Strommenger, B., Altmann, D. and Witte, W. (2009) Nasal colonization of humans with methicillin-resistant *Staphylococcus aureus* (MRSA) CC398 with and without exposure to pigs. PLoS ONE, August: pp e6800.

Danmap 2006, (2007) Use of antimicrobial agents and occurrence of antimicrobial resistance in bacteria from food animals, food and humans in Denmark.

Danmap 2010, (2011) Use of antimicrobial agents and occurrence of antimicrobial resistance in bacteria from food animals, food and humans in Denmark.

Dingle, K.E., Colles, F.M., Ure, R., Wagenaar, J.A., Duim, B., Bolton, F.J., Fox, A.J., Wareing, D.R.A. and Maiden, M.C.J. (2002) Molecular characterization of Campylobacter jejuni clones: a basis for epidemiologic investigation. Emerging Infectious Diseases, 8, 9, 949-955.

EC (2011) European Commission – Communication from the Commission to the European Parliament and the Council – Action plan against the rising threats from Antimicrobial Resistance.

European Council (2012) Council of the European Union – Council conclusions on the impact of antimicrobial resistance in the human health sector and in the veterinary sector – A "One Health" perspective.

EFSA (2009) Scientific report of EFSA – Analysis of the Baseline survey on the prevalence of methicillin-resistant *Staphylococcus aureus* (MRSA) in holdings with breeding pigs, in the EU, 2008. Part A: MRSA prevalence estimates. EFSA Journal, 7, 11: 1376.

EFSA (2010) Scientific report of EFSA – Analysis of the Baseline survey on the prevalence of methicillin-resistant *Staphylococcus aureus* (MRSA) in holdings with breeding pigs, in the EU, 2008. Part B: Factors associated with MRSA contamination of holdings. EFSA Journal, 8, 6: 1597.

EFSA (2011a) European Food Safety Authority, Panel on Biological Hazards (BIOHAZ) – Scientific Opinion on the public health risks of bacterial strains producing extended-spectrum beta-lactamases and/or AmpC beta-lactamases in food and food-producing animals. EFSA Journal, 9, 8, 2322.

EFSA (2011b) Scientific report of EFSA and ECDC – The European Union summary report on trends and sources of zoonoses, zoonotic agents, and food-borne outbreaks in 2009. EFSA Journal, 9, 3, 2090.

EFSA (2011c) Scientific report of EFSA and ECDC – The European Union summary report on antimicrobial resistance in zoonotic and indicator bacteria from humans, animals and food in the European Union in 2009. EFSA Journal, 9, 7, 2154.

Fallacra, D.M., Monahan, C.m., Morishita, T.Y., Bremer, C.A. and Wack, R.F. (2004) Survey of parasites, and bacterial pathogens from free-living waterfowl in zoological settings. Avian Diseases, 48, 759-767.

Foxman, B. (2002) Epidemiology of urinary tract infections: incidence, morbidity and economic costs. American Journal of Medicine, Supplement 1A: 5S-13S.

Hamouda, A., Findlay, J., Hassan, L.A. and Amyes, S.G.B. (2011) Epidemiology of *Acinetobacter baumannii* of animal origin. International Journal of Antimicrobial Agents, 38, 314-318.

HPA (2006) Report Health Protection Agency – *Staphylococcus aureus* bacteraemia: England, Wales and Northern Ireland, January to December 2005.

HPA (2012) Report Health Protection Agency – Quarterly Epidemiological Commentary: Mandatory MRSA, MSSA and *E. coli* Bacteraemia and *C. difficile* Infection Data (up to January –March 2012).

MARAN 2009 (2011) Monitoring of Antimicrobial Resistance and Antibiotic Usage in Animals in the Netherlands in 2009.

Mossad, S.B. (2010) Upper respiratory tract infections. Cleveland Clinic, Center for Continuing Education. <a href="http://www.clevelandclinicmeded.com/medicalpubs/diseasemanagement/infectious-disease/upper-respiratory-tract-infection/">http://www.clevelandclinicmeded.com/medicalpubs/diseasemanagement/infectious-disease/upper-respiratory-tract-infection/</a>

Nijsten, R., London, N., van den Bogaard, A. and Stobberingh, E. (1996) Antibiotic resistance among *Escherichia coli*, isolated from faecal samples of pig farmers and pigs. Journal of Antimicrobial Chemotherapy, 37, 1131-1140.

Prescott, L.M., Harley, J.P. and Klein, D.A. (2005) Chapter 37. The epidemiology of infectious disease and 39. Human diseases caused by bacteria. Microbiology, 6<sup>th</sup> Edition. Eds Prescott, L.M., Harley, J.P. and Klein, D.A. McGraw-Hill, New York, USA.

VMD (2010) Veterinary Medicines Directorate Report – Overview of Antimicrobial Usage and Bacterial Resistance in Selected Human and Animal Pathogens in the UK: 2007.

VLA (2002) Salmonella in livestock production in GB, 2001.

VLA (2007) Salmonella in livestock production in GB, 2006.

Voss, A., Loeffen, F., Bakker, J., Klaasen, C. and Wulf, M. (2005) Methicillin-resistant *Staphylococcus aureus* in pig farming. Emerging and Infectious Diseases, 11, 1965-1966.